



Semen Swim-up Procedure

(Recommended for use when semen has normozoospermic parameters. Usually results in a more variable collection of motile spermatozoa than when using a density gradient).

1. Bring all reagents to 37°C before use. Quinn's[®] Sperm Washing Medium (ART-1005/ART-1006) should be stored at 2° to 8°C until use.
2. Place 1.5 mL of warm Quinn's Sperm Washing Medium in a sterile round bottom culture tube.
3. Using a 3 cc syringe with a 1.5 inch, 21 gauge needle, gently place up to 1 mL of liquefied semen under the medium into the bottom of the tube and cap the tube.
4. Place the tubes in an incubator or water bath at an incline of 30-45° and incubate for one hour. If a CO₂ incubator is used, make sure the tubes are tightly capped.
5. Using a pipette or syringe, carefully remove the upper 1.0 to 1.2 mL of medium down to the interface between the medium and semen. Combine the overlays from several tubes if more than 1 mL of semen was processed.
6. Place the combined overlays into a sterile 15 mL conical centrifuge tube and make the total volume up to 3 mL if necessary with warm Quinn's Sperm Washing Medium.
7. Centrifuge at 500 x g for 5 minutes.
8. Carefully remove the supernatant and resuspend the sperm pellet in a suitable volume of appropriate medium. (e.g. use 0.4 mL of Quinn's Sperm Washing Medium for IUI; use bicarbonate buffered medium such as Quinn's Advantage[®] Fertilization Medium (ART-1020) for IVF and dilute the sperm to an appropriate volume.)

Related Products

Part Number	Description	Unit Size
ART-1005	Quinn's Sperm Washing Medium	12 x 12 mL
ART-1006	Quinn's Sperm Washing Medium	100 mL